

# BxChip Grossing

## Training Video



## Purpose

1. To establish a standard procedure for the accurate and concise grossing of needle-core biopsies placed into the BxChip.

## Scope

1. This procedure applies to laboratories that receive and process needle-core biopsies using the BxChip.

## Definitions

1. Gross examination or “grossing” – the process by which pathology specimens are inspected with the bare eye to obtain descriptive information, while being processed for further microscopic examination.
2. 10% Neutral Buffered Formalin (NBF) – a common general-purpose histology fixative.
3. BxChip<sup>®</sup> – a patented sectionable tissue matrix.
4. BxBoard<sup>®</sup> – a tissue transport and preservation container.

## Reference Documents

1. CAP and AAPA Grossing guidelines.

2. Manual of Surgical Pathology 3<sup>rd</sup> ed.
3. Surgical Pathology Dissection – An Illustrated Guide 2<sup>nd</sup> ed.

## Responsibilities

1. Histology Technicians – to accession cases, verify patient information, and prepare specimens for grossing.
2. Grossing Technicians – to properly identify each specimen for every patient and maintain the specimens' integrity, and accurate cassette labeling at the time of grossing.

## Materials, Supplies, and Equipment

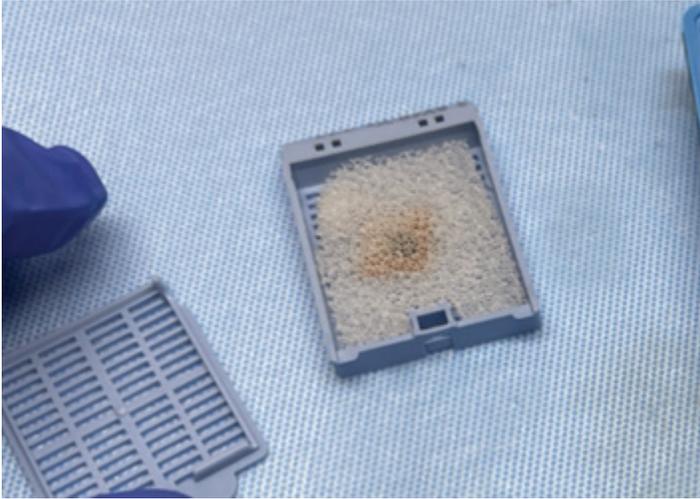
1. Vented hood or grossing station
2. Forceps or tweezers
3. BxChips
4. Labeled cassettes
5. Sponges (contact Lumea for recommendations)
6. Cassette pen/pencil/marker
7. PPE: lab coat, nitrile gloves, eye protection
8. Other lab safety equipment as appropriate

## Safety Concerns

1. Specimen sites and patient information should be verified on the specimen containers and associated documentation.
2. Dispose of used specimen containers in biohazard waste per laboratory protocol.
3. Be cautious of formalin spills and fumes.

## Procedure

1. Open the biopsy kit. Verify patient information and match with any associated paperwork.
2. Obtain cassettes with the appropriate case/accession number.
3. Place a high-flow sponge in the bottom of the cassette.



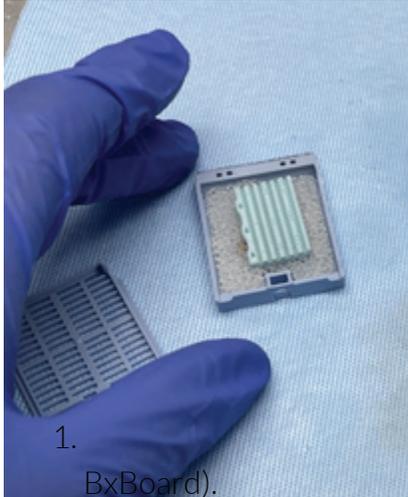
Soak sponges in NBF or saline prior to use. This will minimize tissue sticking to the sponge during processing.

1. Remove a BxChip from the storage container using forceps. Be careful not to tear, puncture, or damage the BxChip.



Do not leave the BxChip out of formalin or saline for more than 15 minutes. It will dry out!

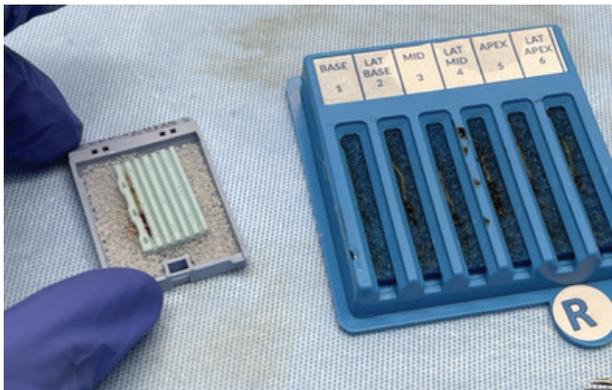
1. Place the BxChip on top of the sponge in the cassette.
  1. Orient and load the BxChip in the same direction each time with the arrows on the left pointing away from the user.



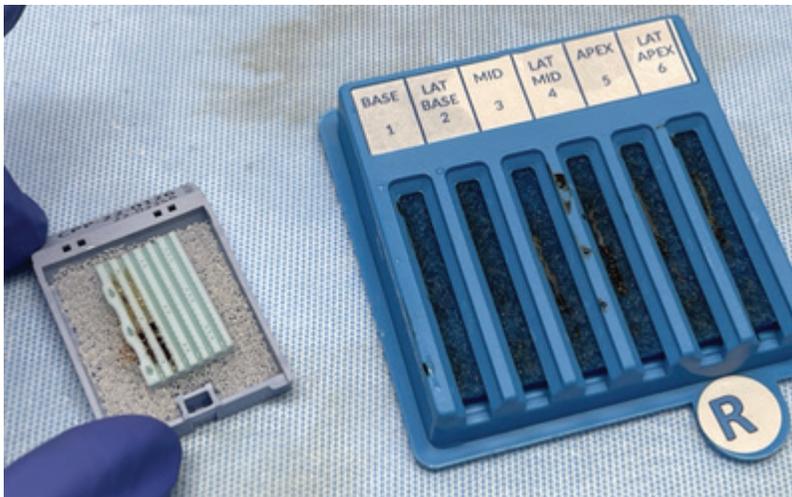
The BxChip is now ready to be loaded with needle core biopsies (shown below transferring from Lumea's

2.

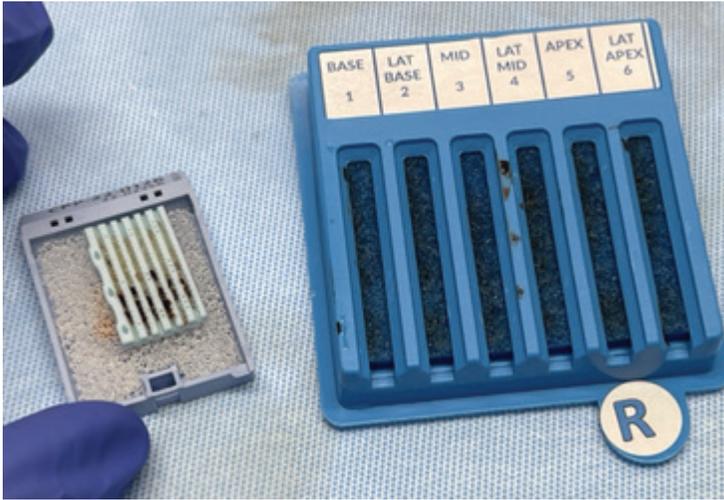
Place the cores in lanes from left to right, one at a time.  
First lane:



1. Second lane:



1. All lanes filled:



1. If using the BxBoard, be sure to maintain the distal and proximal ends of the core as you transfer into the BxChip.

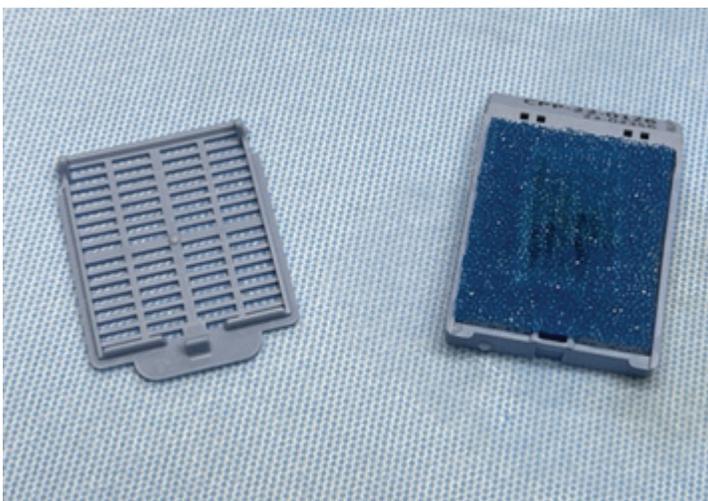
1. If not using the BxBoard, transfer cores from their transport containers one at a time into the subsequent lanes of the BxChip.

1. Document the specimen site names for each lane according to laboratory protocol.

2. Ensure that the Pathologist can positively identify the sites in each BxChip lane for diagnosis.

2. Double check that all the bits of the biopsy cores have been properly placed in the correct lane of the BxChip.

3. Place a wet (NBF or saline) processing sponge on top of the BxChip.



1. Be careful not to slide or drag the sponge across the BxChip as this could disrupt the cores or pull them out of their respective lanes.

2. Place the cassette lid on top of the cassette and lock it into place.



1. Store in NBF or other formalin substitute until processing.
  1. If received on the BxBoard, consider holding in the formalin bath for at least two hours prior to processing or add a fixation step to your processing protocol.
2. Microwave processing tissues can be placed in a 70% Ethanol holding bath for 60 min prior to processing.